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## LIST OF ABBREVIATION

Abbreviation and Symbol	Elaboration
%	Percent
et al.	And his associate
CVASU	Chittagong Veterinary and Animal Sciences University
E.coli	Escherichia coli
MC	MacConkey agar
EMB	Eosin Methylene Blue agar

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The Author

#### ABSTRACT

Avian colibacillosis is considered to be a major bacterial disease in the poultry industry worldwide. The prevalence of colibacillosis in broiler chickens was studied from January to February 2017 that was presented at Department of Pathology and Parasitology, Chittagong Veterinary and Animal Sciences University. 50 swabs from liver, lungs, cloaca of 30 dead birds were collected in sterile nutrient broth as well as 30 liver samples for histopathology. Bacteria were isolated through cultural properties in MacConkey and Eosin Methylene Blue agar. Tissue samples were studied under light microscope after routine Hematoxylline and Eosin staining. Cloudy thickened air sacs, pericarditis, congestion in the liver, lung and spleen were observed in dead birds. On histopathological examination heterophils, lymphocytes, macrophages and fatty liver change found in liver. Based on cultural properties 31 (62%) isolates were found positive. All isolates of Boalkhali were found positive.

Keywords: Colibacillosis, commercial broiler, histopathology, E. coli

#### **CHAPTER-I: INTRODUCTION**

In the world more than a billion people currently live in extreme poverty, these people are powerless, isolated, vulnerable and malnourished (Mack et al., 2004). For this section of people poultry production can be one of the effective tools of combat food security and poverty many parts of the world (Hensen, 2004, Mack et al., 2004). The development and growth of poutry industry over the last 50 years has been one of the remarkable achievements in the history of agriculture. More than 130 hatcheries produce 3.4 million day-old chicks per week and 30,000 commercial broiler and layer farms supply 0.26 million tons of poultry meat and 5210 million eggs per year (Rahman, 2003). With a view to meet the protein gap with shortest possible time, a number of poultry farms have been established on commercial basis in and around the cities and towns and are operated under intensive management. Poultry not only provides protein but also provides full time employment to about 20% of the population and about 50% peoples are associated with this sub sector as part timers. Moreover, poultry farming is a quick returnable enterprise but one of the major constrains in the development of poultry industries is the outbreak of the diseases. Among the different poultry diseases avian Colibacillosis has been found to be a major infectious disease of all ages of birds which causes a variety of disease manifestations in poultry including yolk sac infection, omphalitis, respiratory tract infection, swollen head syndrome, polyserositis, coligranuloma, enteritis, cellulitis and salpingitis. septicemia, Colibacillosis of poultry is characterized in its acute form by septicemia resulting in death and in its subacute form by pericarditis, air sacculitis and peri-hepatitis (Calnek et al., 1997). Yolk sac infection (YSI) is a major cause of mortality of broilers during the first week of post-hatching (Bains, 1979). Broilers suffering from colibacillosis are depressed; show respiratory distress and growth retardation. Mortality usually remains below 5%, but morbidity often reaches more than 50% (Wray et al., 1996; Vandekerckhove et al., 2004). Day Old Chicks may become infected via the yolk sac, but in older chicks the infection is considered to be mainly airborne. Young broiler

chickens up to three weeks of age are highly susceptible to the disease. But colibacillosis commonly occur in three to twelve weeks of age and characterized by intestinal disease. However, various risk factors may increase the susceptibility of broilers to colibacillosis. In addition, the *E. coli* concentration in the air of the broiler house is an important factor. It causes serious loss specially if there is a bad management or stress in broilers such as complicating infections like chronic respiratory disease (CRD) or Mycoplasma (Talha et al., 2003; Rahman, 2003b; Hossain et al., 2004). Heavy loss occurs in broilers due to morbidity, mortality, reduced production and poor chick quality (Islam et al., 2003; Rahman, 2003b; Rahman et al., 2004; Hossain et al., 2004). Investigation on colibacillosis is still scanty in Bangladesh. The disease pattern is variable. This study was undertaken to know the status of colibacillosis in commercial poultry farms.

Chittagong is also a commercial poultry rearing zone of Bangladesh. A significant number of people depend on poultry business. This is important to know the prevalence of colibacillosis in commercial poultry. This study was conducted with the aim to

- 1. Know the prevalence of E. coli in commercial poultry in Chittagong
- 2. Know the pathological changes in different organs due to colibacillosis

## **CHAPTER-II: MATERIALS AND METHODS**

#### Study area and duration

The study was conducted on commercial broiler chicken of various farm of Chittagong, Bangladesh during the period of January to February 2017.

## **Study population**

A total number of 30 dead chickens were brought from various farms of Chittagong to Department of Pathology and Parasitology, CVASU. Sterile swabs were used to take swabs from liver, lungs and cloaca from all 30 chickens.

#### **Study Design**

This study was done by using the following study design:



E.coli positive

## Collection of clinical history about clinical findings

The clinical history was collected from farm owner by cross question method. These questions were asked easy and the scientific and technical terms were avoided.

## Postmortem examination and sample collection

Postmortem examinations of the dead chicken were done by maintaining standard procedures. Sample was collected from liver and yolk sac and by using sterile cotton swab and inoculated into test tube containing Nutrient broth and liver sample kept in Bouin's solution for histopathology.

## Media used for bacterial isolation

MacConkey (MC) agar and Eosin Methylene Blue (EMB) agar were used as solid media. Nutrient broth was used as enrichment of all bacteria. Gram's staining reagents was used for identification of *E. coli*.

Table-1: Methods of culture was done

Sl	Agar	Incubation	time	and	Observations	
		temperature				
1.	MacConkey agar	Incubated	at	$37^{0}$	Dark pink colored raise	d
		temperature fo	or 24 ho	urs.	colony	
2.	Eosin Methylene	Incubated	at	37 <sup>0</sup>	Characteristic metallic sheen	
	Blue (EMB)	temperature fo	or 24 ho	urs		
	agar					

## Data analysis

Data was stored in MS excel (Microsoft Word 2007) and descriptive analysis was done in this study.

## **CHAPTER- III: RESULTS**

### **Results of cultural and biochemical examination**

Among 50 swab samples collected from liver, lung and cloaca of 30 broiler chicken. 70% samples are positive in MacConkey and 62% samples are positive in EMB agar which is also positive in Gram's staining. In the microscopic examination of Gram's staining, all the positive samples are found as Gram-negative, pink colored, rod shaped bacteria which are arranged in single or in pairs.

Table-2: The examination result at a glance done for isolation of E. coli

SL Name of the media /test Total no. No. of positive Percentage of sample sample (%) 1. MacConkey agar 50 35 70 2. Eosin Methylene Blue (EMB) agar 50 31 62 3. Gram 's staining 50 31 62





Figure-1: Prevalence of E. coli in different media

Area	No of sample	EMB Agar (+ve)	Prevalance (%)
Raojan	10	10	100
Darmapur	6	3	50
Vatiari	9	4	44.44
Anowara	5	2	40
Hatazary	7	4	57.14
Betbunia	2	0	0
Lohagara	3	1	33.33
Boalkhali	5	5	100
Abul Khayer steel mill	3	2	66.67

Table-3: Prevalance of E.coli in different sampling sites

Table-3 shows the prevalence of *E. coli* in different sampling sites. Out 50 samples the highest prevalence of was found in Boalkhali (100%), Raojan (100%) and lowest prevalence in Betbunia (0%)



Figure-2: Prevalance of E. coli in different sampling sites

## **Colony characters:**

Pink colonies on MacConkey agar and greenish colonies with metallic sheen on EMB Pink colonies on MacConkey agar and greenish colonies with metallic sheen on EMB agar (Figure-3) after overnight incubation were confirmed as *E. coli* these colony characteristics correspond with the finding others.

## **Staining characters:**

Microscopy revealed Gram-negative, pink ,short rod-shaped organism arranged singly or pairs (Figure-4).

## **Pathological study**

Postmortem findings revealed cloudy and thickened air sacs (air sacculitis), congested liver and consolidate lung in some chickens. The pericardium was thickened and the spleen enlarged with congestion. All these lesions indicated the septicaemic form of colibacillosis. E. coli can cause several disease conditions but in the present study not all conditions induced by E.coli were recorded.

## Microscopic lesions:

The liver showed coagulation type of focal necrosis, infiltration of heterophils, lymphocytes and macrophages mainly in portal area and fatty change (Figure-5).



Figure- 5: Section of liver with <i>E. coli</i> infection showing fatty change	Figure-6: Section of liver with <i>E.coli</i> infection showing Mononuclear cell infiltration

#### **CHAPTER-IV: DISCUSSION**

Avian Colibacillosis caused by E. coli is a major health problem in poultry industry. The study was conducted with the aim of isolation and identification of Escherichia coli present in liver and yolk sac of dead chicken. In this study, chicken clinically infected with colibacillosis manifested characteristics clinical sign such as watery diarrhea, anorexia, weakness and loss of body weight. A similar type of clinical sign of colibacillosis was also recorded (Kim et al., 1996). Here this study observed that 62 E. coli was present in broiler Chicken. The results was partially agreed with the findings of Maiorka et al. (2006) where he reported that at hatching, chick was still microorganismfree but after hatching a significant number of microorganisms invaded and colonized in different organ. In this study, the average finding of E. coli was 62% from commercial broiler which is similar to the earlier reports of Suha et al. (2008) who reported 43.50%. Rahman et al. (2004) reported 67.73% and Ahmed et al., (2009) found 52.26% colibacillosis in commercial broiler which is higher than this study. The result is also similar with Bhattacharjee et al., (1996) reported 40.82% of E. coli in chicken from Bangladesh. Morphology, staining and cultural characteristics of the bacteria in different cultural media as recorded in the study were almost similar as reported by Choudhury et al. (1985). They reported that staining and morphology of isolated E. coli exhibited Gram negative, small rod, arranged in single or pairs, non-spore former.

## **CHAPTER-V: CONCLUSION**

Avian colibacillosis caused by *E. coli* is a major health problem in a poultry industry. Although unrespective sample size was investigated in this study but based on the findings it can be assumed that a higher population of chickens in our country is suffering from colibacillosis. The poor sanitation and handling of sewage and poor feeding, poor housing overall poor hygienic management and vaccination failure could be a source contamination. Colibacillosis can be treated if appropriate antibiotics are chosen to treat the disease.

#### **CHAPTER-VI: REFERENCES**

- Ahmed MS, Sarker A and Rahman MM (2009). Prevalence o f infectious diseases of broiler chickens in Gazipur district, Bangladesh. *Journal o f Veterinary Medicine*, 23(2): 326-331.
- Hossain M.K, Ahmed M, Kabir H, Sarker MRR, Jalil MA, Adhikary GN 2004:Poultry diseases at Rajshahi in Bangladesh. Journal of Animal and Veterinary Advances 3 657-659.
- Islam MR, Das BC, Hossain KH, Lucky, NS, Mostafa MG 2003: A study on the occurrence of poultry disease in Sylhet Region of Bangladesh. International Journal of Poultry Science 2 354-356.
- Rahman M 2003a: Growth of poultry industry in Bangladesh: Poverty alleviation and employment. In: Proceedings of 3<sup>rd</sup> International Poultry Show and Seminar, from February 28 to March 2, 2003, held in Bangladesh China frindship Conference Center (BCFCC) Sher-e Bangle Nagar, Dhaka Bangladesh pp: 1-7.
- Rahman MA 2003b: Disease profile with pathogenicity and drug sensitivity of salmonellosis, colibacillosis and pasteurellosis in commercial chickens. MS Thesis. Department of Medicine, Bangladesh Agriculture University, Mymensingh pp.71.
- Talha AFSM, Hossain MM, Chowdhury EH, Bari ASM, Islam MR, Das PM 2001: Poultry diseases occurring in Mymensingh district of Bangladesh. Bangladesh Veterinary Journal 18 20-23.
- Kim, H.J., Kong, M.I., Clung, U.K., Kim, H.J., Kang, M.I. and Clung, U.I. (1996) Survey of enteric diseases in Chickens. Korean J. Vet. Res., 36: 1007-1012.
- Biswas PK, Faruque R, Ahmed S and Dey VC (2001). Antibiotic Sensitivity Pattern of Pathogenic Escherichia coli isolated from Fayoumi chicken. *Bangladesh Journal of Microbiology*, 18(2): 121-126.
- Bhattacharjee PS, Kundu RL, Biswas PK, Mazumder JU, Hossain E and Miah AH (1996). A retrospective analysis of chicken diseases diagnosed at the Central

Disease Investigation Laboratory, Dhaka. *Bangladesh Veterinary Journal*, 30 (3–4): 105–113.

- Cretikos M, Telfer B, McAnulty J (2008). Enteric disease outbreak reporting, New South Wales, Australia, 2000 to 2005. *New South Wales Public Health Bullettin*, 19 (1-2): 3-7.
- Castanon JI (2007). History of the use of antibiotic as growth promoters in European poultry feeds. *Poultry Science*, 86: 2466–2471.
- Calnek BW (1997). Diseases of Poutry, 10<sup>th</sup> Edition, Iowa State University Press. Ames, Iowa, USA. pp: 83-89, 131, 151-153.
- Choudhury KA, Amin MM, Rahman A and Ali MR (1985). Investigation of natural outbreak of fowl cholera. *Bangladesh Veterinary Journal*, 19(1-4): 49-56.
- Davies J (1994). Inactivation of antibiotics and the dissemination of resistance genes. *Science*, 264: 375-382.
- Guerra B, Junker E, Schroeter A, Malorny B, Lehmann S, Helmuth R (2003). Phenotypic and genotypic characterization of antimicrobial resistance in German *Escherichia coli* isolates from cattle, swine and poultry. *Journal of Antimicrobial and Chemotherapy*, 52 (3): 489-92.
- Jawetz E, Melnick J and Adelberg EA (1984). Review of Medical Microbiology. 16th ed. Los Altos, California: Long Medical Publication, pp. 122-144.
- Lu J, Sanchez S, Hofacre C, Maurer JJ, Harmon BG and Lee MD (2003). Evaluation of broiler litter with reference to the microbial composition as assessed by using 16S rRNA and functional gene markers. *Applied Environmental Microbiology*, 69: 901–908.
- Maiorka A, Dahlke F and Maria MS (2006). Broiler adaptation to post hatching period. *Ciencia Rural, Santa Maria*, 36: 65-71.
- NCCLS (2009). Performance standards for antimicrobial disk susceptibility tests. Approved standard M2-A6. Wayne, Pa: National Committee for Clinical Laboratory Standards.

Neu HC (1992). The crisis in antibiotic resistance. Science, 257: 1064-1073.

- Osterblad M, Hakanen A, Manninen R, Leistevuo T, Peltonen R, Meurman O, Huovinen P and Kotilainen P (2000). A between-species comparison of antimicrobial resistance in enterobacteria in fecal flora. *Antimicrobial Agents and Chemotherapy*, 44: 1479-1484.
- Rahman MA, Samad MA, Rahman MB and Kabir SML (2004). Bacterio-pathological studies on Salmonellosis, Colibacillosis and Pasteurellosis in natural and experimental infections in chicken. *Bangladesh Journal of Veterinary Medicine*, 2(1): 1-8
- Suha A, Husseina ALIH, Hassanb RI and Sulaima NC (2008). Bacteriological and pathological study of yolk sac infection in broiler chicks in sulaimani district. *Journal of Dohuk University*, 11(1): 124-127.
- Tankson JD, Thaxton JP and Vizzier TY (2002). Bacteria in heart and lungs of young chicks. *Journal of Applied Microbiology*, 92: 443-450.
- Van de Boogard AE, Stobberingh EE (2000). Epidemiology of resistance to antibiotics links between animals and humans. *International Journal Antimicrobial Agents*, 14: 327-335
- URL : Bacterial diseases of poultry: Http <u>www.msstate.edu/dept/poultry/disbact</u>. htm. Int.
- Wray C, Davies RH and Corkisch JD (1996). Enterobacteriaceae. In: Jordan FTW and Pattison M (Eds.). Poultry Diseases. WB Saunders, Cambridge, UK. pp. 9-43.

# APPENDIX

Formula of various bacteriological media

A. MacConkey Agar	
Peptone	
Lactose	
Bile salts	5.0g
Sodium chloride	5.0g
Neutral red	0.075g
Agar	12.0g
Distilled water	

B. Eosin Methyl Blue (EMB) Agar

10.0g
5.0g
5.0g
2.0g
13.5g
0.4g
0.065g
1000ml

### LIMITATIONS

Sample size of this investigation was not representative to the population due to short period of the study.

Timing was not sufficient for such type of study.

Follow up was not completed for each and every case.

Lack of my subject related journals to help the work.

### **Diagnostic information's**

Case no:

Date.....

1. Name of the farm:

Basic information's:

- 2. Name of the owner:
- 3. Address of the farm:
- 4. Type of farm: (layer/Broiler).
- 5. Housing system: (intensive/semi intensive)
- 6. Floor type:(Litter/Concentrate/Slat/Mud/Others)

7. a) Vaccination: 1=Properly, 2=Improperly b) Vaccination: (Good/Moderate/Not performed).

8. a) Biosecurity: Visitor access: 1=Not restricted, 2=Restricted. Foot bath: 1=Present,
2= Absent Wild birds/animal access: 1=Yes, 2=No. Chick care management: 1=
Properly 2=Not properly.

- 9. Age of bird:....
- 10. Flock size:....
- 11. Morbidity.....
- 12. Mortality.....

Clinical findings:		
A. General systemic status		
1	2	.3
B. Respiratory system:		
1	2	3
C. Digestive system:		
1	.2	3
D. External organ:		
1	2	3

# Information about clinical findings:

A. General systemic status:

Clinical findings	Nature/additional information
Dullness/Depression	Colisepticemic form
Dehydration	Due to enteritis
Ruffled feathers	Colisepticemic form
Weakness	Coligranuloma form

# B. Respiratory system:

Clinical findings	Nature/additional information
Gasping/open beck breathing	In case of pericarditis due to colibacillosis and gasping followed by death.
Respiratory distress	In colisepticemia form due to bronchopneumonia

# C. Digestive system:

Clinical findings	Nature/additional information
Diarrhoea	Due to enteritis and yellow colour faeces in young
Anorexia	In later stage of swollen head syndrom
Reduce feeding	Colisepticemic form

# D. External organ:

Clinical findings	Nature/additional form
Pasty vent	In Colisepticemia form, due to steaky dropping
Swollen and inflamed naval region	In case of yolk sac infection, due to oedema in naval region.
Swollen joint	In case of infective arthritis due to septicaemia and synovitis. Here mainly hock joint is mainly affected.
Omphalitis	In case of yolk sac infection.

## BIOGRAPHY

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