## **Methane Emission From Sheep**



#### A PRODUCTION REPORT SUBMITTED BY

Niaj Ahmed Shimul Intern ID. 08 Roll No:16/08 Reg No: 1613 Session: 2015-2016

A production report submitted in partial satisfaction of the requirements for the degree of Doctor of Veterinary Medicine

#### (DVM)

**Faculty of Veterinary Medicine** 

CHATTOGRAM VETERINARY AND ANIMAL SCIENCES UNIVERSITY

Zakir Hossain Road, Khulshi, Chattogram-4225

# **Methane Emission From Sheep**



Signature of the Author Intern ID. 08 Roll No: 16/08 Reg. No: 1613 Session: 2015-2016

# Faculty of Veterinary Medicine CHATTOGRAM VETERINARY AND ANIMAL SCIENCES UNIVERSITY

Zakir Hossain Road, Khulshi, Chattogram-4225

## **Methane Emission From Sheep**



Signature of Supervisor **Professor Ashraf Ali Biswas Director of Research and Extension** Department of Animal Science And Nutrition. Faculty of Veterinary Medicine

# Faculty of Veterinary Medicine CHATTOGRAM VETERINARY AND ANIMAL SCIENCES UNIVERSITY

Zakir Hossain Road, Khulshi, Chattogram-4225

### Table of contents

Content	Page
Abstract	IV
Chapter 1: Introduction	1-6
Chapter 2: Materials and Methods	7-8
Chapter 3: Results and Discussion	9
Limitations	10
Conclusion	11
References	12-15
Acknowledgement	16
Biography	17

### List of Table

Table	Title	Page
Table 1. H	Enteric methane emission from different livestock species in	n the world6
Table 2. A	Amount of methane detected at different time from sheep	9

### Abstract

Ruminant livestock can produce 250 to 500 L of methane per day. Many factors influence methane emissions from ruminant and include the following: level of feed intake, type of carbohydrate in the diet, feed processing, addition of lipids or ionophores to the diet, and alterations in the ruminal micro flora. On a global scale agriculture and in particular enteric fermentation in ruminants is reported to produce about one fourth (21 to 25%) of the total anthropogenic emissions of methane (CH<sub>4</sub>). Methane is produced during the anaerobic fermentation of hydrolyzed dietary carbohydrates in the rumen and represents an energy loss to the host besides contributing to emissions of greenhouse gases into the environment. However, there appears to be uncertainty in the CH<sub>4</sub> estimation from livestock due to the limited availability of data to document the variability at the farm level and also due to the significant impact of diet on the enteric CH<sub>4</sub> production. The methane mitigation strategies require robust prediction of emissions from rumen. There are many methods available which would be suitable for measuring CH<sub>4</sub> produced from the various stages of animal production. However, several factors need to be considered in order to select the most appropriate technique like the cost, level of accuracy required and the scale and design of the experiments to be undertaken. We collected methane gas in polythene bag with bottle and estimated amount of gas by Gas Chromatography. Result shows highest emission 984.84 PPM at 1 pm after feeding and lowest emission 17.29 PPM at 9:30 am before feeding.

Keywords: Ruminant, Methane, Emission, Gas chromatography.

### **Chapter 1: Introduction**

Methane ( $CH_4$ ) is a colorless, odorless gas produced as a byproduct of ruminant animals' gastrointestinal tract microbial fermentation of feed. The term "methanogen" refers to bacteria that produce methane. Methanogens create methane from hydrogen and carbon dioxide. As a result of microbial fermentation, hydrogen and carbondyoxide are generated. Enteric fermentation in ruminant animals produces methane, a powerful gas.

Methane (CH<sub>4</sub>) has been linked to ozone depletion in the stratosphere (Blake and Rowland, 1988). When CH<sub>4</sub> is oxidized, water vapour is released into the stratosphere, which could provide surfaces for heterogeneous processes that degrade ozone. Global initiatives like the Kyoto Protocol demand that these emissions be minimized or at the very least avoided from increasing further (Howden and Reyenga, 1999). With a 100-year global warming potential (GWP) 23 times that of CO<sub>2</sub>, methane is the second most significant contributor to global warming (IPCC, 2001). Despite being present in the atmosphere at much lower concentrations than CO<sub>2</sub>, CH<sub>4</sub> is said to be responsible for roughly 20% of the greenhouse gas effect (IPCC, 1990; 1992).

 $CH_4$  is produced largely by ruminant eructation from microbial fermentation of hydrolyzed dietary carbohydrates such as cellulose, hemicellulose, pectin, and starch in the rumen. Hydrogen and  $CO_2$  are the key substrates for ruminal methanogenesis. The majority of hydrogen created during the fermentation of hydrolyzed dietary carbohydrates, particularly during the conversion of hexose to acetate or butyrate, ends up in CH<sub>4</sub>. Microbial fermentation of amino acids, which produces ammonia, volatile fatty acids,  $CO_2$ , and  $CH_4$ , can also produce significant amounts of  $CH_4$ . Methane causes a large energy loss in ruminant animals, accounting for around 8% of gross energy at maintenance intake levels and declining to about 6% as intake levels rise. Increased knowledge and quantification of  $CH_4$  generation in the rumen has implications for both global environmental protection and effective animal production.

It is critical to have faith in the accuracy of the CH<sub>4</sub> measurement equipment in order to create an accurate inventory or apply mitigation techniques. Methane emissions from livestock have been evaluated as part of research into ruminal fermentation, energy balance, feed additive evaluation, and most recently, to describe and minimize ruminant contribution to the global  $CH_4$  load. Respiration calorimetry equipment such as complete body chambers, head boxes, ventilated hoods, and face masks have been used to monitor livestock CH<sub>4</sub> emissions (Johnson and Johnson, 1995). The data gathered through these methods served as the basis for the prediction equations that were utilized to create mathematical models and national and global inventories (Benchaar et al., 1998; Mills et al., 2001). The precise estimation of CH<sub>4</sub> emissions from ruminants under a variety of conditions is critical for developing measures to reduce CH<sub>4</sub> emissions. There are a variety of methods that may be used to measure CH<sub>4</sub> generated at various phases of animal production. They are ECD-Electron capture detector; FID-Flame ionization detector; FTIR-Fourier transform infrared (spectroscopy); GC-Gas chromatography/Gas chromatograph; TCD-Thermal conductivity detector; TDL-Tuneable diode laser; TGA-Trace gas analyzer; SF6- Sulfur hexafluoride. However, various criteria must be considered in order to choose the best technique, including the cost, the level of precision necessary, and the scale and design of the experiments to be conducted (Johnson et al., 2000).

#### **Objectives:**

- 1. To Know about gas produced by sheep.
- 2. To Determine suitable methane detection method from sheep.

#### **Review of the literature**

Sheep are grazing animals, and the forages they graze have an effect on intestinal CH<sub>4</sub> output. Because it is difficult to identify the exact feed intake, CH<sub>4</sub> emissions from sheep on grazing pasture are difficult to forecast; the nutritional value of the pasture may fluctuate both between and within seasons. In general, the nutritious content of forages is determined by their maturation level. The nutritious content of forages diminishes as pasture matures, while the concentration of structural carbohydrates rises, decreasing pasture's nutritional value. As a result, when sheep feed on low-quality pastures, acetic fermentation will prevail in the rumen, increasing CH<sub>4</sub> output. The amount of CH<sub>4</sub> produced by comparatively nutrient-rich pasture will be lower than that produced by poor-quality pasture (Clark *et al.*, 2011). Sheep are grazing animals in general, but due to the intensification of the production system, they are now reared in many regions of the world under semi-intensive and intensive feeding systems. The amount of enteric CH<sub>4</sub> produced in the rumen of sheep is determined by the feed consumed, the substrate degraded, and the type of end products created. Because the synthesis of propionate requires H<sub>2</sub>, whereas the formation of acetate and butyrate releases H<sub>2</sub>, rumen fermentation that produces more propionate lowers enteric CH<sub>4</sub>. As a result, any feeding regimen that encourages rumen fermentation to shift from acetate to propionate would reduce H<sub>2</sub> release and CH<sub>4</sub> output (Basarab et al., 2013), and these associations have been demonstrated in many of the studies in sheep.

The chamber method has good accuracy and precision for assessing the daily production of  $CH_4$  from housed animals but limited capacity with regards to the number of animals (Storm *et al.*, 2012).

Though the adoption of a methodology for the estimation of  $CH_4$  depends on many factors, in vivo techniques such as GreenFeed, sulfur hexafluoride tracer technique and respiration chambers are instrumental in order to estimate the precise emission and could be useful in determining the national  $CH_4$  emission when a large number of experiments are conducted involving large animals and locally available seasonal feedstuffs with repeated measurements.

Polythene tunnels are just like respiration chambers, but these are easy to transport and operate and can be placed on pasture where animals are grazing. These tunnels overcome the limitation of respiration chambers where measurement of enteric  $CH_4$ emission in pasture-grazing sheep is difficult. Polytunnels are used for measuring the emission in grazing animals without much disturbance to their natural behavior (Malik *et al.*, 2017).

Methane emissions increased ( $P \le 0.05$ ) with increasing live weight, feeding level measured as multiples of maintenance and digestibility of dry matter and decreased for rations with wider ratios of crude fibre intake and intake of N-free extracts. Crude fibre content in the ration and energy density of the ration showed no clearly identifiable effect on methane emissions (pelchen and peters 1998).

CH<sub>4</sub> production from the open-circuit chambers was greater than from the tunnel system (Murray *et al.*, 1999).

Enteric CH<sub>4</sub> contributes 17% and 3.3% of the global CH<sub>4</sub> and greenhouse gases (GHGs) emissions, respectively (Knapp *et al.*, 2014).

Regional estimates by various agencies suggested a huge disparity in enteric  $CH_4$  emission across the globe, and countries like Latin America, Africa, China and India hold first, second, third and fourth position in enteric  $CH_4$  emission, respectively (Malik *et al.*, 2016).

Facemask also uses the same principle as in chamber and hood for quantifying the CH<sub>4</sub> emission from livestock (Liang *et al.*, 1989).

Laser methane detector (LMD) could be a quick and reliable method for measuring  $CH_4$  emission from sheep in a stress-free natural environment (Chagunda *et al.*, 2009).

Infrared Thermography technique relies on the principle that the animal's body surface temperature is related to the feed efficiency which in turn affects the degree of  $CH_4$  emission from the animal. Montanholi et al. (2008)

The intraruminal gas measurement device was developed by the Commonwealth Scientific and Research Organization in Australia to estimate enteric  $CH_4$  emission from ruminants. The device can be used as an alternative to SF6 tracer technique and respiration chambers. The device is impermeable to liquid and is placed into the stomach of the animal. The animals swallow the device as a bolus with tubular body.

The bolus is permeable to gases and has gas sensors that can detect  $CH_4$  in rumen. A controller is attached to the gas sensor so that it can periodically process and give out data regarding the amount of  $CH_4$  in the rumen (Wright *et al.*, 2013).

In vitro gas production test is employed in laboratory where conditions akin to the rumen are simulated in an artificial environment and gas production is recorded, which is subsequently analyzed on gas chromatograph for  $CH_4$ . This technique is a powerful tool for generating real-time data in short duration for a large number of samples for their  $CH_4$  reduction abilities (Lee *et al.*, 2003).

Portable accumulation chambers (PAC) may be used for screening a large number of animals in order to select low-CH<sub>4</sub>-emitting sheep. PAC allows you to test a huge number of animals in a short amount of time. Pastoral Greenhouse Gas Consortium (PGgRC) in New Zealand is developing one such PAC, in which sheep walking into the chamber will be held within for roughly 25–30 minutes while the CH<sub>4</sub> content is measured. A translucent polycarbonate box makes up the PAC. When the sheep is in the polycarbonate box, the concentration of CH<sub>4</sub> in the box rises, giving an estimate of CH<sub>4</sub> emission. A gas detector is used to track the change in CH<sub>4</sub> content in the chamber's environment over time. The results of this technique were comparable to those of respiratory chamber measurements (Goopy *et al.*, 2009, 2011, 2016).

All the ruminant species emit  $CH_4$  by virtue of their digestive system which is adapted for anaerobic fermentation, especially the rumen, the largest of the four chambers. Most ruminant species in underdeveloped nations rely on low-quality roughages to meet their nutritional needs, and as a result, they contribute significantly to global  $CH_4$  emissions.

Livestock species	<sup>a</sup> Enteric CH <sub>4</sub> emission (kg $\times 10^9$ )
Cattle	69.9
Buffalo	10.7
Sheep	6.04
Goat	4.61
Swine	1.08
Camel	1.11
Horse	1.05
Ass	0.42
Mule	0.11
Alpaca	0.063
Total $CH_4$ production	94.9

Table 1. Enteric methane emission from different livestock species in the world

<sup>a</sup>Estimated methane emitted by different livestock species in 2010. Adopted from Patra (2014a)

Cattle and buffaloes, for example, produce more CH<sub>4</sub> than smaller ruminants (Table 1). According to one estimate, total global enteric CH<sub>4</sub> emission (kg×10<sup>9</sup>) from various animal classes will reach 105 by 2025, with cattle, buffalo, sheep, goat, swine, camel, horse, ass, mule, and alpaca contributing 77.3, 12.1, 6.18, and 5.19, 1.29, 1.17, 1.03, 0.45, 0.09, and 0.13, respectively (Patra, 2014a). Sheep and goats, which are smaller ruminants, produce somewhat more CH<sub>4</sub> than horses and swine, which are non-ruminants. The amount of CH<sub>4</sub> emitted by different ruminants is determined by a variety of parameters such as the animal's species, population, dry matter intake, degree of production, pasture quality, roughage quality, rumen volume, and other factors (Broucek, 2014).Sheep and goats, on the other hand, can create 10–16 kg CH<sub>4</sub> per year, whereas cattle can produce 60–160 kg per year, depending on the animal's size and dry matter consumption. Non-ruminant herbivore animals such as donkeys, horses, mules, and others produce CH<sub>4</sub> as a result of anaerobic fermentation in their hindgut.

### **Chapter 2: Materials and Method**

#### **Materials required**

Rubber band, water bottle, polythene bag, three way cannula, syringe, vacutainer tube, Gas chromatography machine.

**Gas Chromatography:** The principle is based on the individual partitioning characteristics of various gases in the sample between a mobile phase (such as Helium) and a stationary solid phase packed in a column. Each component was identified by its retention time on the column and quantified by a subsequent detector after the components in the gaseous mixture were separated. The detector is the most important component of the GC system. For measuring greenhouse gases, three types of detectors are typically used: thermal conductivity detectors (TCD) for CO<sub>2</sub>, flame ionization detectors (FID) for CH<sub>4</sub>, and electron capture detectors (ECD) for N<sub>2</sub>O.The detectors can be connected to GC systems singly or in groups, allowing for the simultaneous investigation of multiple gases (Sitaula *et al.*, 1992). CH<sub>4</sub> has detection limits of less than 200 ppb, according to Crill *et al.*, (1995).

#### Procedure

Bottom part of a water bottle was cutted circularly. A round rubber band was attached with the cutted part of the bottle. The open part (the bottom) was inserted into the mouth of the sheep. As it takes air, one valve (upper position) becomes open, but when it releases air, this valve is closed and that gas will go through the mouth of the bottle to the collection bag (polythene). Collection of methane was done from sheep for 4 days at different times of the day: 9:30 am, 1 pm, 5:30 pm, and 6:10 pm.

Gas was then collected in a vacutainer tube with the help of a syringe and a 3-way cannula. Here we needed restraining of the animal properly. As the animal's mouth was inserted through the facemask, it was hard to make the animal calm and quiet. Gas was collected for 5 minutes, 10 minutes, and 15 minutes. Expelled gas from the

mouth was collected. After collecting the gas, it was transferred into a vacutainer tube. Here, caution was taken so that the gas would not be contaminated.

We tried to separate methane from  $CO_2$ . Sample was inserted in container carrying NaOH. Indicator was added with NaOH.  $CO_2$  will react with NaOH to form Na<sub>2</sub>CO<sub>3</sub>. With this conversion the color of the solution will be changed. But in our case no color change occurred. So separation of  $CO_2$  was not possible here from the sample.

### **Chapter 3: Results and Discussion**

The experiment shows that methane released by sheep was higher after feeding the animal concentrate and roughage at 1 pm. Methane concentration can vary with animal size, feed intake, genetic variation etc.

Time	9:30 am	1 pm	5:30 pm	6:10 pm
	32.6465	959.027	376.808	69.558
Methane concentration	47.996	984.847	374.619	100.037
(PPM)				
	17.297	782.368	310.414	39.079
Average Methane	32.6465	908.7473	353.947	69.558
concentration (PPM)				

Table 2. Amount of methane detected at different time from sheep

Concentrate feed was given to the sheep at 12 pm. Sheep get roughage from pasture land. Each sheep get 250 gm of gram as concentrate. As the table 2 is showing that average methane concentration is highest at 1 pm (908.743 ppm). At 5:30 pm the average concentration of the methane decreased to 353.947 ppm. And lastly At 6:10 pm methane emission was declining more. But in the morning methane concentration was lowest because they were in empty stomach at that time.

# Limitations

As methane collection device was not fully accurate, there was a chance of contamination with other atmospheric gas. And we were able to detect limited amount of methane from sheep.

# Conclusion

The amount of methane detected was very less amount. But the experiment shows the relationship between the feed intake and methane production. Methane production was high after feeding and less before feeding.

### References

- Blake, D. R. and F. S. Rowland. 1988. Continuing worldwide increase in tropospheric methane, 1978 to 1987. Sci. 239:1129-1131.
- Howden, S. M. and P. J. Reyenga. 1999. Methane emissions from Australian livestock: implications of the Kyoto Protocol. Aust. J. Agric. Res. 50:1285-1291.
- Johnson, K. A. and D. E. Johnson. 1995. Methane emissions from cattle. J. Anim. Sci. 73:2483-2492.
- Benchaar, C., J. Rivest, C. Pomar and J. Chiquette. 1998. Prediction of methane production from dairy cows using existing mechanistic models and regression equations. J. Anim. Sci. 76: 671-672.
- Mills, J. A. N., J. Dijkstra, A. Bannick, S. B. Camel, E. Krebreab and J. France. 2001. A mechanistic model of whole-tract digestion and methanogenesis in the lactating dairy cow: model development, evaluation and application. J. Anim. Sci. 79:1584-1597.
- Johnson, D. E., K. A. Johnson, G. M. Ward and M. E. Branine. 2000. Ruminants and other animals. In: Atmospheric Methane: Its role in the global environment, (Ed. M. A. K. Khalil), Springer-Verlag, Berlin Heidelberg, pp. 112-133.
- Sitaula, B. K., J. Luo and L. R. Bakken. 1992. Rapid analysis of climate gases by wide bore capillary gas chromatography. J. Environ. Quality. 21:493-496.
- Crill, P. M., J. H. Butler, D. J. Cooper and P. C. Novelli. 1995. Standard analytical methods for measuring trace gases in the environment. In: (Ed. P. A. Matson

and R. C. Harriss): Biogenic trace gases: Measuring emissions from soil and water. Methods in ecology series. Oxford, Blackwell Sci. pp. 164-205.

- Storm IMLD, Hellwing ALF, Nielsen NI, Madsen J (2012) Methods for measuring and estimating methane emission from ruminants. Animals 2:160–183
- Malik, Pradeep Kumar "Methane Estimation Methodologies in Sheep." *Sheep Production Adapting to Climate Change*. Springer, Singapore, 2017. 267-290.
- Murray PJ, Moss A, Lockyer DR, Jarvis SC (1999) A comparison of systems for measuring methane emissions from sheep. J Agric Sci 133:439–444
- Knapp J, Laur G, Vadas P (2014) Invited review: enteric methane in dairy cattle production: quantifying the opportunities and impact of reducing emissions. J Dairy Sci 97:3231–3261
- Malik PK, Kolte AP, Dhali A (2016) GHG emissions from livestock: challenges and ameliorative measures to counter adversity. In: Manning AJ (ed) Greenhouse gases-selected case studies. Intech, Winchester
- Liang JB, Terada F, Hamaguchi I (1989) Efficacy of using the face mask technique for the estimation of daily heat production of cattle. In: Van Der Honing Y, Close WH (eds) Energy metabolism of farm animals. Pudoc, Waginingen, pp 348–351
- Chagunda M, Ross D, Roberts D (2009) On the use of a laser methane detector in dairy cows. Comput Electron Agric 68:157–160
- Montanholi YR, Odongo NE, Swanson KC (2008) Application of infrared thermography as an indicator of heat and methane production and its use in

the study of skin temperature in response to physiological events in dairy cattle (*Bos taurus*). J Therm Biol 33:468–475

- Wright ADG, Ellis K, Dempsey J, (2013) System, method and device for measuring a gas in the stomach of a mammal. World intellectual property organization. Patent WO 2013/003892 AI
- Lee HJ, Lee SC, Kim JD (2003) Methane production potential of feed ingredients as measured by in vitro gas test. Asian Australas J Anim Sci 16:1143–1150
- IPCC. 1990. Climate Change: The IPCC Impact Assessment. Canberra, Australia: Australian Government Publishing Service.
- IPCC. 1992. Climate Change 1992. The Supplementary Report to the Scientific Assessment (Ed. J. T. Houghton, B. A. Callander and S. K. Varney). Cambridge: Cambridge University Press.
- IPCC. 2001. Climate Change 2001. The Scientific Basis. Contribution of working group I to the Third Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge Press.
- Goopy JP, Hegarty RS, Robinson DL (2009) Two hour chamber measurement provides useful estimate of daily methane production in sheep. In: Chilliard Y, Glasser F, Faulconnier Y, Bocquier F, Veissier I, Doreau M (eds) Ruminant physiology: digestion, metabolism, and effects of nutrition on reproduction and welfare. Wageningen Academic Publishers, Wageningen, pp 190–191
- Goopy JP, Woodgate R, Donaldson A et al (2011) Validation of a short-term methane measurement using portable static chambers to estimate daily methane production in sheep. Anim Feed Sci Technol 166-167:219–226

- Goopy JP, Robinson DL, Woodgate RT et al (2016) Estimates of repeatability and heritability of methane production in sheep using portable accumulation chambers. Anim Prod Sci 56:116
- Clark H, Kelliher F, Pinares-Patiño C (2011) Reducing CH4 emissions from grazing ruminants in New Zealand: challenges and opportunities. Asian Australas J Anim Sci 24:295–302
- Basarab JA, Beauchemin KA, Baron VS et al (2013) Reducing GHG emissions through genetic improvement for feed efficiency: effects on economically important traits and enteric methane production. Animal 7:303–315
- Patra AK (2014a) Trends and projected estimates of GHG emissions from Indian livestock in comparisons with GHG emissions from world and developing countries. Asian-Aust J Anim Sci 27:592–599
- Broucek J (2014) Production of methane emissions from ruminant husbandry: a review. J Environ Prot 5:1482–1493

## Acknowledgement

All praises are due to Almighty "Allah" who has created everything of the nature and who enable me to complete this study. I feel great pleasure to express my deepest sense of gratitude and indebtedness to my supervisor Ashraf Ali Biswas, Professor, department of Animal Science and Nutrition, Chattogram Veterinary and Animal Sciences University for his scholastic guidance, valuable suggestions, constant inspiration and encouragement throughout the entire period of my study. Special thanks to other teachers and staff, Department of Animal Science and Animal

Nutrition, for their valuable advice and co-operation.

# **Biography**

I am Niaj Ahmed Shimul, son of Atiqul Islam Babul and Shamima Yasmin. I Passed my Secondary School Certificate examination from Gafargaon Islamia Govt. High School, Gafargaon in 2013 and Higher Secodery Certificate examination from Shahid Syed Najrul Islam College, Mymensingh in 2015. I enrolled for Doctor of Veterinary Medicine (DVM) degree in Chattogram Veterinary and Animal Sciences University (CVASU), Bangladesh. In future, I want to be a veterinary practitioner and want to contribute to the development of the nation.