

**INNOVATION OF LOW COST METHANE CONCENTRATION
ESTIMATION METHOD & COMPARING METHANE
CONCENTRATION OF SHEEP & GOAT**



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ABSTRACT

Ruminants are one of the main contributors to the release of greenhouse gases into the atmosphere. Large ruminants are capable of producing 250 to 500 Liters of methane (CH₄) daily. CH₄ is a by-product of feed fermentation that is released primarily through eructation by ruminants. Methanogenesis is carried out by anaerobic organisms called Archaea which are found in the rumen where the anaerobic conditions are optimal. There prevails several methods or technique to estimate the concentration of CH₄ eructed by the ruminants. Most of the techniques are too costly and requires highly skilled personnel. We used sensor based hand-held gas detector to detect the CH₄ concentration in exhaled air of sheep and goat. It is easy to operate and less expensive. We performed the experiment for two days at CVASU animal farm and recorded the PPM reading given by the gas detector. The data clearly demonstrate that CH₄ concentration in exhaled air of sheep and goat increase after feeding and reaches at the highest level in between 1.5 to 2 hours after feeding. 1 hour after feeding, significant variation between CH₄ concentration of sheep and goat was found in both days. The Mean CH₄ concentration of goats at 11:00 AM were 1077.42 PPM (1st day) and 1009 PPM (2nd day) which were higher than the mean CH₄ concentration of sheep 908.66 PPM (2nd day) and 856.96 PPM (2nd day).

Keyword: Methane, Fermentation, Gas detector, Sheep and Goat.

CHAPTER-I: INTRODUCTION

Methane is the important factor of global warming. It was reported that approximately 20% of greenhouse effect is due to methane (CH₄) (IPCC, 1990; 1992). Depletion of stratospheric Ozone is promoted by CH₄ (Blake & Rowland, 1988). Methane emission is responsible for approximately 3-14% loss in gross energy intake (Hellwing et al. 2016). So it is important to understand the global methane production and its sources.

Methane production by ruminant is one of the largest sources of global warming. Approximately 15% of global methane emissions are due to ruminants (Gerber et al. 2013). Ruminants yield methane by anaerobic fermentation of feed as by-product & emitted primarily by eructation (Janssen PH et al. 2008). About 87% of methane released from the animals mouth and nose originates from the forestomach through eructation and blood absorption (Murray et al. 1976). About 13% of the CH₄ is originated in the hindgut, where 89% is absorbed into the circulatory system and excreted via expiration (Ricci et al., 2014). The organisms responsible for methanogenesis are known as Archaea which located at the rumen that gives them ideal anaerobic condition. The methanogenic bacteria use two biochemical pathways to yield methane gas, such as the reduction of Carbon dioxide (CO₂) and decarboxylation of acetate. The reduction of CO₂ by Hydrogen (H₂) is the major one. The pattern of the methane production by ruminants is pulsatile, with high concentration of methane in exhaled breath coinciding with each eructation.

Open-circuit respiration chamber (Waghorn, 2014), Sulfur hexafluoride tracer technique (SF₆, Johnson et al., 2007) & the automated heat-chamber system (GreenFeed, Zimmerman & Zimmerman, 2012) are the most accepted methods for estimation of CH₄. Open-circuit breathing chambers for whole animals are the most popular right now, and they come in all shapes and sizes, from simple poly tunnels and shower curtains over stalls (Powell et al., 2007; Aguerre et al., 2011), to more complex and pricy dedicated calorimeters that require more time and money to set up (Global Research Alliance, 2012). These methods are too costly & require highly skilled labor which limits their utilization in developing country like ours. Some methods which are less accurate such as Laser based Methane estimation Device (LMD) is explored to measure CH₄ concentration in air emitted by the animals (Chagunda et al. 2009).

Recent research disclosed that there is variation in CH₄ production between individual animals when fed identical feed. Also the CH₄ emission rate is not constant & may vary time to time. Feed additives play an important part in rumen fermentation and CH₄ emission (Seon-Ho Kim et al., 2020). Consumption of lysozyme in the form of dietary supplements has been demonstrated to enhance rumen fermentation in vitro and to decrease CH₄ emissions (Ashraf A. Biswas et al., 2016). Genetic factors that affect the feeding behavior, live weight, age, type of feed etc are the possible factors for the variation.

The main objectives of this study are:

- To detect the methane concentration in exhaled air of goat & sheep.
- To determine the variation of concentration of methane in exhaled air before & after feeding.
- To establish a low cost methane concentration estimation method.

CHAPTER-II: MATERIALS & METHODS

To perform this experiment, we used SPD203 Methane gas detector. It can detect the gas and show the percentage concentration on digital display. This uses highly sensitive sensor for accurate measurement & detection.

The experiment was conducted in CVASU animal farm under the Department of Animal Science & Nutrition, Chattogram Veterinary & Animal Sciences University, Khulshi, Chattogram, Bangladesh.

To conduct the experiment, we formed two groups of 4 goats and sheep each. Body weight of all the animals was measured via electric weight machine.

Table-1: Body weight of sheep

Animal Id.	Body weight (Kg)
S-10	21.12
S-20	20.43
S-30	19.75
S-40	21.81

Table-2: Body weight of goat

Animal Id.	Body weight (Kg)
G-11	13.72
G-20	17.16
G-30	17.5
G-40	20.36

Each animal was given 400 grams of the same type of concentrate feed separately at 10:00 AM. Readings were taken from each animal in gas detector machine half an hour before giving feed. After that, readings were taken from each animal at intervals of half an hour after feeding until the readings reached the same level. We perform this activity two days in a row.

In this way, we measured and recorded the concentration of methane gas in the exhaled air of each goat and sheep respectively. And the recorded data was analyzed statistically by STATA software to compare methane concentration between sheep and goat.

CHAPTER-III: RESULTS AND DISCUSSION

On the basis of the data collected, it can be concluded that in both sheep and goat, the methane concentration in the breath air is low prior to consumption and increases post-consumption. Within 1.5 to 2 hours following ingestion, the methane concentration in the exhaled air reaches its highest level in both species. Subsequently, the methane concentration decreases gradually.

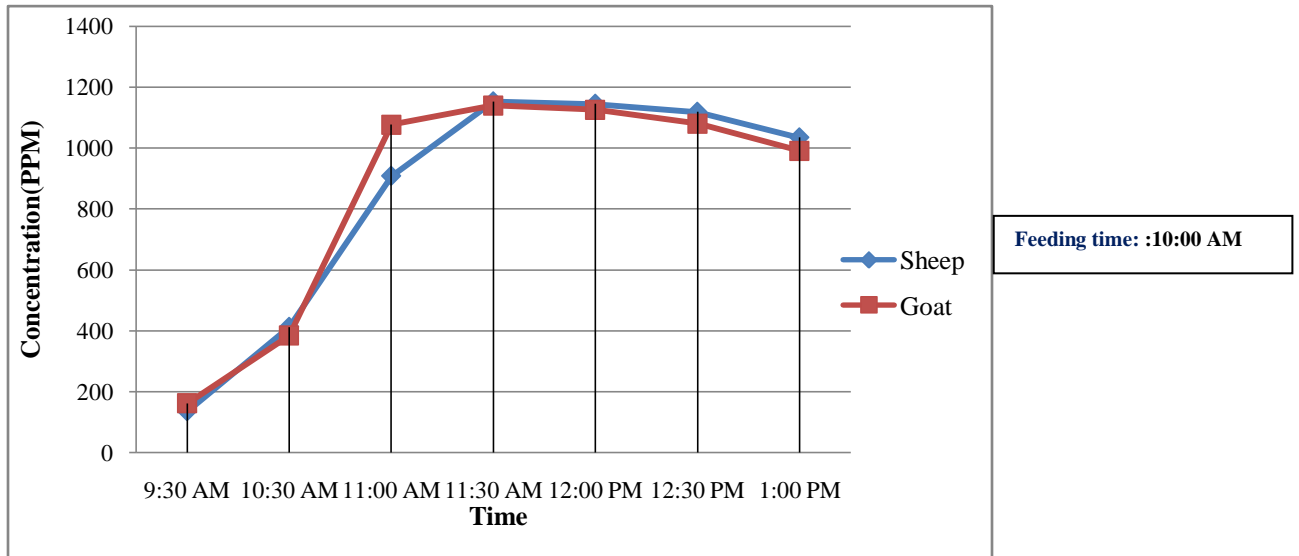


Fig-1: Methane emission concentration(PPM) of exhaled air on day-1

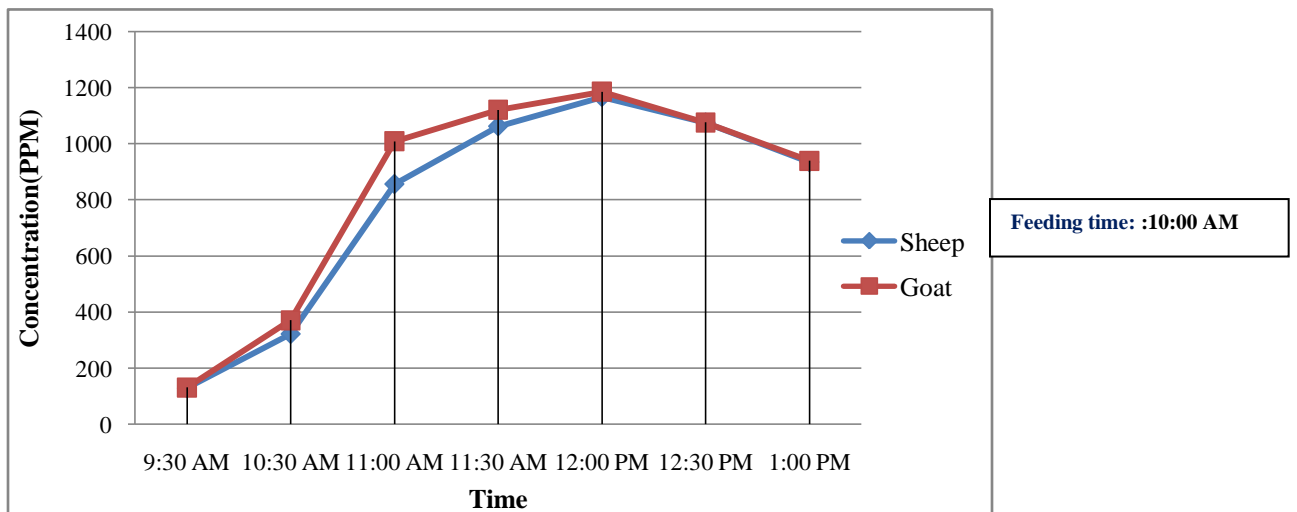


Fig-2: Methane emission concentration(PPM) of exhaled air on day-2

Table 3: Methane emission concentration Day 1

Time	Mean \pm SD		SEM		T value	P value
	Sheep	Goat	Sheep	Goat		
9:30 AM	135.92 \pm 6.88	162.25 \pm 27.08	3.44	13.54	1.89	0.107
10:30 AM	412.83 \pm 22.74	384.75 \pm 52.74	11.37	26.37	0.98	0.365
11:00 AM	908.66 \pm 42.59	1077.42 \pm 33.51	21.29	16.75	6.23	0.001
11:30 AM	1153.25 \pm 18.88	1140 \pm 18.05	9.44	9.03	1.01	0.351
12:00 PM	1145.33 \pm 35.43	1125.75 \pm 33.85	17.71	16.93	0.799	0.455
12:30 PM	1119 \pm 28.47	1080.75 \pm 40.48	14.23	20.24	1.546	0.173
1:00 PM	1034.96 \pm 42.82	990.54 \pm 13.49	21.41	6.745	1.98	0.106

SD= Standard Deviation; SEM= Standard Error of Mean

Table 4: Methane emission concentration Day 2

Time	Mean \pm SD		SEM		T value	P value
	Sheep	Goat	Sheep	Goat		
9:30 AM	129.08 \pm 7.56	131.41 \pm 28.65	3.78	14.32	0.16	0.878
10:30AM	322.37 \pm 49.18	371.71 \pm 53.53	24.59	26.765	1.36	0.223
11:00AM	856.96 \pm 40.78	1009 \pm 27.46	20.39	13.73	6.18	0.001
11:30AM	1063.08 \pm 42.75	1121.5 \pm 45.1	21.37	22.55	1.88	0.109
12:00PM	1167.5 \pm 49.03	1186.75 \pm 62.38	24.52	31.19	0.485	0.644
12:30PM	1076.17 \pm 54.12	1076.92 \pm 33.46	27.06	16.73	0.024	0.982
1:00 PM	935.25 \pm 44.69	940.33 \pm 42.37	22.34	21.18	0.165	0.874

SD= Standard Deviation; SEM= Standard Error of Mean

There was no significant difference ($P > 0.05$) between the CH₄ concentration of sheep and goat before feeding. After feeding, in case of both 1st and 2nd day at 10:30 AM, 11:30 AM, 12:00 PM, 12:30 PM and 1:00 PM, we found no significant difference ($P > 0.05$) of methane concentration in exhaled air of sheep and goat. But 1 hour interval after feeding at 11:00 AM we found significant difference ($P < 0.05$) between CH₄ concentration of sheep and goat in both days.

The mean CH₄ concentration of goats were 1077.42 PPM (1st day) and 1009 PPM (2nd day) which is higher than the mean CH₄ concentration of sheep 908.66 PPM (2nd day) and 856.96 PPM (2nd day). The fig-3 indicates also a considerable difference among sheep (882.205 PPM) and goats (1041.335PPM) at 11:00 AM in the total mean CH₄ concentration.

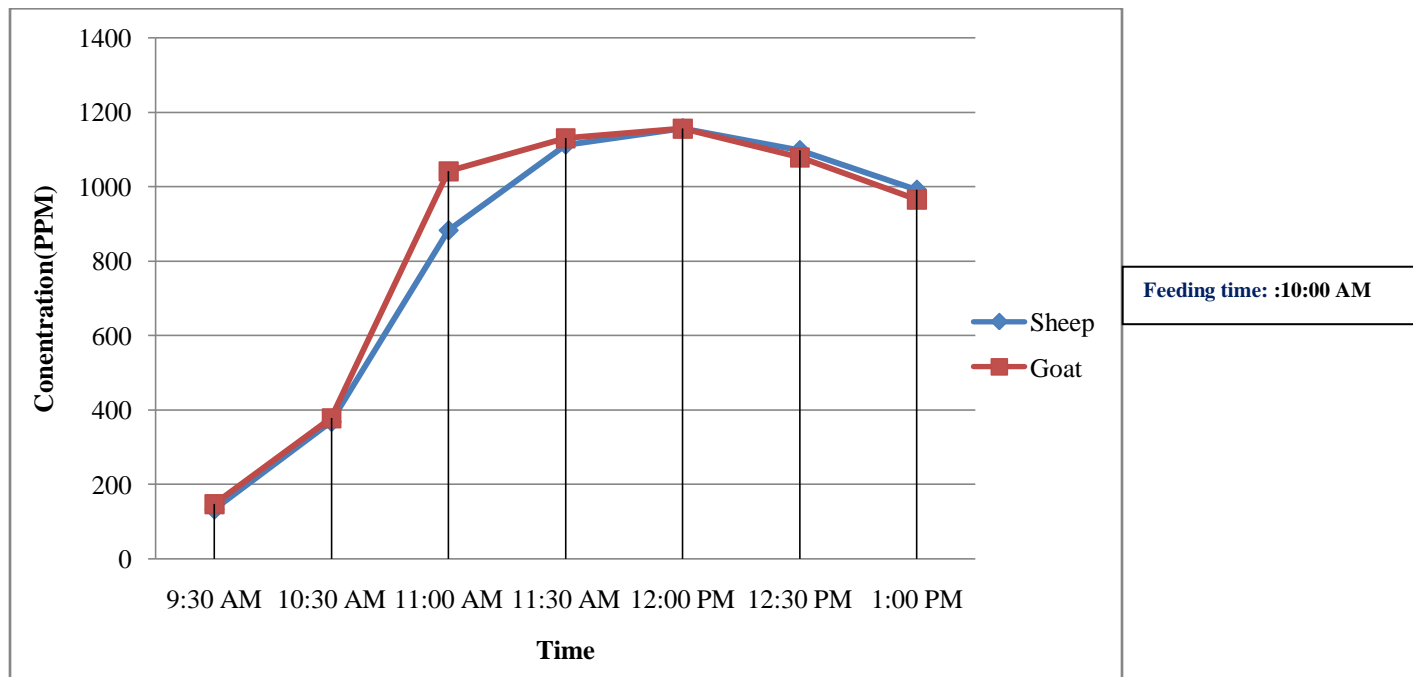


Fig-3: Total mean methane emission concentration(PPM) of exhaled air

Methane concentration variation between sheep and goats can be attributed to differences in their digestive systems. Both animals are ruminants, but their diets and digestive processes vary. Sheep typically consume more fibrous plants, leading to a slower fermentation process in their stomachs and thus lower methane production. Goats, on the other hand, have more diverse diet that includes a broader range of plants, leading to faster fermentation and potentially higher methane emissions. Other factors like body weight, gut microbiota composition and metabolic differences can also contribute to the variation in methane concentrations between these two animals.

CHAPTER-IV: CONCLUSIONS

The results demonstrate that the CH₄ concentration values prior to feeding did not vary significantly between the sheep and goat. There is also no significant difference in sheep and goat for the same type and the same amount of feed consumed in a given period of interval. But in between 1 to 1.5 hour interval after feeding there is significant variation in methane concentration of sheep and goat. Confidently, more correct values could be obtained if the concentration of CH₄ measured in more precise way and the data were compared using a most recognized and accepted method. However, it's important to consider factors such as calibration, sensor accuracy, and maintenance to ensure reliable results.

CHAPTER-V: REFERENCES

- Aguerre, M.J., Wattiaux, M.A., Powell, J.M., Broderick, G.A., Arndt, C., 2011. Effect of forage-to-concentrate ratio in dairy cow diets on emission of methane, carbon dioxide, and ammonia, lactation performance, and manure excretion. *J. Dairy Sci.* 94:3081-3093.
- Blake, D. R. and F. S. Rowland. 1988. Continuing worldwide increase in tropospheric methane, 1978 to 1987. *Sci.* 239:1129- 1131.
- Biswas, Ashraf A., et al. "Use of lysozyme as a feed additive on in vitro rumen fermentation and methane emission." *Asian-Australasian journal of animal sciences* 29.11 (2016): 1601
- Chagunda M, Ross D, Roberts D (2009) On the use of a laser methane detector in dairy cows. *Comput Electron Agric* 68:157–160
- Gerber, P. J., Steinfeld, H. & Henderson, B. (2013) Tackling climate change through livestock: a global assessment of emissions and mitigation opportunities.
- Greenfeed, Zimmerman PR, Zimmerman RS (2012) Method and system for monitoring and reducing ruminant CH₄ production.
- Global Research Alliance, 2012. Technical Manual on Respiration Chamber Design, In: Pinares-Patiño, C.S., Waghorn, G. (Eds.), Ministry of Agriculture and Forestry, New Zealand. <http://www.globalresearchalliance.org/wp-content/uploads/2012/03/GRAMANFacility-BestPract-2012-FINAL.pdf>. Accessed 12th January 2014.
- Hellwing, A.L.F., Weisbjerg, M.R., Brask, M., Alstrup, L., Johansen, M., Hymøller, L., & Lund, P. (2016). Prediction of the CH₄ conversion factor (Y_m) for dairy cows on the basis of national farm data. *Animal Production Science*
- Janssen PH, Kirs M. Structure of the archaeal community of the rumen. *Appl. Environ. Microbiol.* 2008;74:3619–3625. doi: 10.1128/AEM.02812-07.

- Kim, S. H., Islam, M., Biswas, A. A., Cho, K. K., & Lee, S. S. (2020). Effects of Detoxified Sulfur as a Feed Supplement on in Vitro Rumen Fermentation and Methane Mitigation. *생명과학회지*, 30(9), 743-748.
- Murray, R.M., Bryant, A.M., Leng, R.A., 1976. Rates of production of methane in the rumen and large intestine of sheep. *Brit. J. Nutr.* 36, 1-14.
- Powell, J.M., Cusick, P.R., Misselbrook T.H., Holmes, B.J., 2007. Design and calibration of chambers for measuring ammonia emissions from tie-stall dairy barns. *Trans. ASABE* 49:1139–1149.
- Ricci, P., Chagunda, M.G.G., Rooke, J., Houdijk, J.G., Duthie, C.-A., Hyslop, J., Roehe, R., Waterhouse, A., 2014. Evaluation of the laser methane detector to estimate methane emissions from ewes and steers. *J. Anim. Sci.* 92, 5239-5250.
- Johnson KA, Westberg HH, Michal JJ, Cossalman MW (2007). The SF₆ tracer technique: CH₄ measurement from ruminants. In 'Measuring CH₄ production from ruminants'. (Eds HPS Makkar, PEVercoe) pp. 33-67. (Springer: Dordrecht, The Netherlands).
- Waghorn, G. (2014). Technical Manual on Respiration Chamber Designs.

CHAPTER-VI: LITERATURE REVIEW

Methanogenic bacteria under anaerobic conditions form methane in the rumen of ruminants. This process allows ruminants to use energy in low calorie feed such as grass and fodder that has a high content of cellulose. The level of feed intake and animal's digestibility are determining factors for methane emissions from ruminants (Blaxter et al. 1965). Feed additives play an important part in rumen fermentation and CH₄ emission (Seon-Ho Kim et al., 2020). Consumption of lysozyme in the form of dietary supplements has been demonstrated to enhance rumen fermentation in vitro and to decrease CH₄ emissions (Ashraf A. Biswas et al., 2016). For calculating emissions at a specific level the relation that they have identified is used.

Since 1950, there has been an increase in the number of domesticated animals in the world. Due to an increase in number of ruminants and growing milk production, GHG emissions from ruminants are increasing. Between 1966 and 1986, the methane emissions from domestic animals were 0.6 Tg/year or 0.75 per year (Crutzen et al. 1986). Lerner et al. (1988) compiled a global database of methane emissions from livestock and found that only five countries account for half of global emissions: India, Brazil, The former Soviet Union, USA and China. For 2010, Kelliher and Clark (2010) estimated that methane emissions from ruminants would amount to approximately 100 Tg per year, in which 14 Tg from China, 12 Tg from Brazil, 11 Tg from India, 5.5 Tg from the USA, 3 Tg from Australia, 3 Tg from Pakistan, 3 Tg from Argentina, 2.5 Tg from Russia, 2.3 Tg from Mexico and 2 Tg from Ethiopia. According to a study by Das et al., (2020), livestock GHG emissions from enteric fermentation in Bangladesh were estimated to be 30,124 gigagram.

In animals, there is no universally superior method for CH₄ quantification. Some methods are better suited to small scale scenarios, mainly due to the fact that they have been developed with this in mind and others were built from scratch as a result of their conception for wide scale use. However, for the measurement of enteric methane emissions from ruminants is carried out with a wide range of techniques and methods. In the past ten years, a number of novel CH₄ measurement techniques have been developed which can be applied to a wide range of animals and are suitable for use on farms (Hammond et al., 2016, Biswas et al., 2019).

The respiration chamber is a well established and documented as a reliable CH₄ measurement system (Hammond et al. 2011). Open-circuit breathing chambers for whole animals are the most popular right now, and they come in all shapes and sizes, from simple poly tunnels and shower curtains over stalls (Powell et al., 2007; Aguerre et al., 2011), to more complex and pricy dedicated calorimeters that require more time and money to set up (Global Research Alliance, 2012). An animal is held in a containment chamber large enough for them to be comfortably accommodated, and kept under somewhat negative atmospheric pressure. This prevents any foreseeable gaseous leaks coming in instead of out (Johnson et al. 1995; storm et al. 2012).

Zimmerman (1993) developed the SF₆ technique, and Johnson et al. (1994) were the first to report use of this method for estimating ruminant CH₄ emissions. This method is suitable for confined or free range and grazing animals, in which a permeation tube with the known rate of SF₆ gas leakage into the reticulorumen has been placed inside the animal. In general, average CH₄ emissions are provided by the SF₆ technique. For a given animal, it may differ from that obtained in respiration chambers. Despite the fact that a mean CH₄ emission cannot differ, within and the SF₆ technique used in sheep has considerably increased variation between animals (Pinares-Patino et al., 2011) and dairy cattle (Grainger et al., 2007) relative to the respiration chamber technique.

For the purpose of obtaining CH₄ emission measurements over a 24 hour period, the use of respiratory chambers and SF₆ technique is generally employed. But short term measurement methods offer the ability to detect the presence of CH₄ at specific time points. These methods are typically automated, minimally invasive and non-tempering, thus enabling a high animal throughput. Repeated short-term measurements using facemasks have been used in previous studies (Washburn and brody, 1937).

The GreenFeed system uses a static, short term measurement device to measure CH₄ emissions from individual animals by combining air flow, gas level and head position detection during each animals stay in the unit (Huhtanen et al., 2015). In this system, the collected air is blended, filtered and the airflow is measured by means of an anemometer with hot film coating. The CH₄ concentration in the sample is determined by non-disperse infrared analysis.

Estimation of methane emissions from the animal is based on the background corrected ratio of CH₄ to CO₂ and the predicted CO₂ emissions, as provided by Madsen et al. (2010). The precision of the CH₄ emission estimation based on the CH₄ to CO₂ ratio will be contingent upon a number of variables, including the origin of gases present in air sample, as well as the diurnal variations in the CH₄:CO₂ ratio due to variations in animal behavior and fermentation rate related to meal size and feeding frequency (Madsen et al., 2010).

A measurement of CH₄ concentration in air eructed from cattle at milking has been reported for the first time by Garnsworthy et al., 2012. A sample inlet is inserted into the feed collector of an automated milking system. The gas concentration in the air in the feed manger are taken at 1 second intervals, analyzed and recorded. The method excludes CH₄ in the inhaled air from all eructation's, as well as any flatulence (Garnsworthy et al., 2012). The fact that the distance between the head of the animal and the point of sampling has a significant influence on CH₄ and CO₂ concentration, which is not a factor in the overall air sampling, is a concern for the techniques used to measure gas concentrations in exhaled air samples (Hegarty, 2013).

The use of handheld laser detectors for measuring CH₄ concentration in air between animal's nose or mouth and LMD is a further option to monitor the exhaled level of CH₄ (Ricci et al., 2014). The analysis only relies on peak values reflecting CH₄ concentration increases due to inhalation or eructation. The average values of CH₄ concentration can be readily provided by the LMD. It has the potential to generate accurate estimation of methane from ruminants, thus making it a useful tool for on-field monitoring and decision making for GHG mitigation measures (Chagunda et al., 2013).

REFERENCES

- Aguerre, M.J., Wattiaux, M.A., Powell, J.M., Broderick, G.A., Arndt, C., 2011. Effect of forage-to-concentrate ratio in dairy cow diets on emission of methane, carbon dioxide, and ammonia, lactation performance, and manure excretion. *J. Dairy Sci.* 94:3081-3093.
- Blake, D. R. and F. S. Rowland. 1988. Continuing worldwide increase in tropospheric methane, 1978 to 1987. *Sci.* 239:1129- 1131.
- Biswas, Ashraf A., et al. "Use of lysozyme as a feed additive on in vitro rumen fermentation and methane emission." *Asian-Australasian journal of animal sciences* 29.11 (2016): 1601.
- Biswas, A., and A. Jonker. "Circadian variation in methane emissions by cattle and correlation with level and composition of ryegrass-based pasture eaten." *NZJ Anim. Sci. Prod* 79 (2019): 61-64.
- Blaxter, K L and Clapperton, J L. 1965. Prediction of the amount of methane produced by ruminants. *Br J Nutr*, 19: 511–522. [Crossref], [PubMed], [Web of Science ®]
- Das NG, Sarkar NR and Haque MN (2020). An estimation of greenhouse gas emission from livestock in Bangladesh. *Journal of advance veterinary and animal research*, 7(1): 133-140.
- M. G. G. Chagunda, D. Ross, J. Rooke, T. Yan, J.-L. Douglas, L. Poret, N. R. McEwan, P. Teeranavattanakul & D. J. Roberts (2013) Measurement of enteric methane from ruminants using a hand-held laser methane detector, *Acta Agriculturae Scandinavica, Section A — Animal Science*, 63:2, 68-75, DOI: [10.1080/09064702.2013.797487](https://doi.org/10.1080/09064702.2013.797487)
- Grainger, C., Clarke, T., McGinn, S.M., Auldist, M.J., Beauchemin, K.A., Hannah, M.C., Waghorn, G.C., Clark, H., Eckard, R.J., 2007. Methane emissions from dairy cows measured using the sulfur hexafluoride (SF₆) tracer and chamber techniques. *J. Dairy Sci.* 90, 2755-2766.

Global Research Alliance, 2012. Technical Manual on Respiration Chamber Design, In:Pinares-Patiño, C.S., Waghorn, G. (Eds.), Ministry of Agriculture and Forestry, New Zealand.<http://www.globalresearchalliance.org/wp-content/uploads/2012/03/GRA-MANFacility-BestPract-2012-FINAL.pdf>. Accessed 12th January 2014.

Garnsworthy, P.C., Craigon, J., Hernandez-Medrano, J.H., Saunders, N., 2012. On-farm methane measurements during milking correlate with total methane production by individual dairy cows. *J. Dairy Sci.* 95, 3166-3180.

Huhtanen, P., Cabezas-Garcia, E.H., Utsumi, S., Zimmerman, S., 2015. Comparison of methods to determine methane emissions from dairy cows in farm conditions. *J. Dairy Sci.*98, 3394-3409.

Hammond KJ, Crompton LA, Bannink A, Dijkstra J, Yáñez-Ruiz DR, O’Kiely P, Kebreab E, Eugène MA, Yu Z, Shingfield KJ, Schwarm A, Hristov AN, Reynolds CK 2016. Review of current in vivo measurement techniques for quantifying enteric methane emission from ruminants. *Animal Feed Science and Technology* 219: 13-30.

Hammond, K.J.; Hoskin, S.O.; Burke, J.L.; Waghorn, G.C.; Koolaard, J.P.; Muetzel, S. Effects of feeding fresh white clover (*Trifolium repens*) or perennial ryegrass (*Lolium perenne*) on enteric CH₄ emissions from sheep. *Anim. Feed Sci. Technol.* 2011, 166– 167, 398–404. [CrossRef]

Hegarty, R.S., 2013. Applicability of short-term emission measurements for on-farm quantification of enteric methane. *Anim.* 7, s2, 401-408.

J. Lerner et. Al (1988)Methane emission from animals: A Global High-Resolution Data Base.

Johnson, K. A., & Johnson, D. E. (1995). CH₄ emissions from cattle. *Journal of animal science*, 73(8), 2483-2492.

- Johnson, K.A., Huyler, M., Westberg, H., Lamb, B., Zimmerman, P., 1994. Measurement of methane emissions from ruminant livestock using a SF₆ tracer technique. *Environ. Sci.Technol.* 28, 359-362.
- Kim, S. H., Islam, M., Biswas, A. A., Cho, K. K., & Lee, S. S. (2020). Effects of Detoxified Sulfur as a Feed Supplement on in Vitro Rumen Fermentation and Methane Mitigation. *생명과학회지*, 30(9), 743-748.
- Kelliher, F M and Clark, H. 2010. “Ruminants: Chapter 9”. In Methane and climate change, Edited by: Reay, D, Smith, P and van Amstel, A. 136–150 London: Earthscan.
- Madsen, J., Bjerg, B.S., Hvelplund, T., Weisbjerg, M.R., Lund, P., 2010. Methane and carbon dioxide ratio in excreted air for quantification of the methane production from ruminants. *Livest. Sci.* 129, 223-227.
- Powell, J.M., Cusick, P.R., Misselbrook T.H., Holmes, B.J., 2007. Design and calibration of chambers for measuring ammonia emissions from tie-stall dairy barns. *Trans. ASABE* 49:1139–1149
- Paul J. Crutzen et al. (1986) Methane production by domestic animals, wild ruminants, other herbivorous fauna, and humans. Pages 271-284
- Pinares-Patiño, C.S., Lassey, K.R., Martin, R.J., Molano, G., Fernandez, M., MacLean, S., Sandoval, E., Luo, D., Clark, H., 2011. Assessment of the sulphur hexafluoride (SF₆) tracer technique using respiration chambers for estimation of methane emissions from sheep. *Anim.Feed Sci. Technol.* 166-167, 201-209.
- Ricci, P., Chagunda, M.G.G., Rooke, J., Houdijk, J.G., Duthie, C.-A., Hyslop, J., Roehe, R., Waterhouse, A., 2014. Evaluation of the laser methane detector to estimate methane emissions from ewes and steers. *J. Anim. Sci.* 92, 5239-5250.

Storm, I.M.L.D.; Hellwing, A.L.F.; Nielsen, N.I.; Madsen, J. Methods for measuring and estimating CH₄ emission from ruminants. *Animals* 2012, 2, 160–183.

Washburn, L.E., Brody, S., 1937. Growth and development XLII. Methane, hydrogen, and carbon dioxide production in the digestive tract of ruminants in relation to the respiratory exchange, In: Mumford, F.B. (Ed.), *Growth and Development*, University of Missouri, Columbia, Missouri.

Zimmerman, P.R., 1993. System for measuring metabolic gas emissions from animals. United States Patent number US005265618A.

CHAPTER-VII: APPENDIX

Table 5: Methane emission concentration Day 1(Sheep)

	Before feeding	After feeding					
Time	9:30Am	10:30AM	11:00AM	11:30AM	12:00PM	12:30PM	1:00PM
	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)
Sheep-10	139.33	379.67	851.33	1168.00	1174.67	1144.33	1083.33
Sheep-20	125.67	421.00	931.00	1169.00	1172.33	1139.67	1058.67
Sheep-30	138.33	419.33	903.33	1129.67	1134.33	1108.33	999.50
Sheep-40	140.33	431.33	949.00	1146.33	1100.00	1083.67	998.33

Feeding time: 10:00 AM

Table 6: Methane emission concentration Day 1(Goat)

	Before feeding	After feeding					
Time	9:30Am	10:30AM	11:00AM	11:30AM	12:00PM	12:30PM	1:00PM
	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)
Goat-11	171.67	411.33	1049.33	1127.33	1123.00	1039.67	971.67
Goat-20	131.00	374.67	1053.67	1145.67	1153.67	1113.67	1001.00
Goat-30	194.33	315.67	1085.00	1124.00	1147.33	1117.33	999.50
Goat-40	152.00	437.33	1121.67	1163.00	1079.00	1052.33	990.00

Feeding time: 10:00 AM

Table 7: Methane emission concentration Day 2(Sheep)

	Before feeding	After feeding					
Time	9:30Am	10:30AM	11:00AM	11:30AM	12:00PM	12:30PM	1:00PM
	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)
Sheep-10	131.67	257.33	803.50	1087.50	1202.00	1126.33	984.50
Sheep-20	118.33	352.00	855.33	1103.33	1214.33	1100.00	961.00
Sheep-30	130.33	312.50	867.00	1006.50	1143.00	1077.67	904.50
Sheep-40	136.00	367.67	902.00	1055.00	1110.67	1000.67	891.00

Feeding time: 10:00 AM**Table 8:** Methane emission concentration Day 2(Goat)

	Before feeding	After feeding					
Time	9:30Am	10:30AM	11:00AM	11:30AM	12:00PM	12:30PM	1:00PM
	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)	Avg. (PPM)
Goat-11	112.33	378.67	992.00	1056.33	1233.33	1080.00	946.00
Goat-20	102.00	374.50	982.50	1156.67	1175.33	1086.67	912.00
Goat-30	161.00	301.67	1018.00	1127.33	1235.33	1110.33	998.00
Goat-40	150.33	432.00	1043.50	1145.67	1103.00	1030.67	905.33

Feeding time: 10:00 AM

Statistical Analysis

```
. ttest zerohour , by (Animal)
Two-sample t test with equal variances
```

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	135.915	3.439316	6.878631	124.9696	146.8604
1	4	162.25	13.53849	27.07698	119.1645	205.3355
combined	8	149.0825	8.159678	23.07905	129.7879	168.3771
diff		-26.335	13.96852		-60.51474	7.844736

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.0542
Ha: diff != 0
Pr(|T| > |t|) = 0.1084
Ha: diff > 0
Pr(T > t) = 0.9458
t = -1.8853
degrees of freedom = 6

```

```
. ttest firsthour , by (Animal)
Two-sample t test with equal variances
```

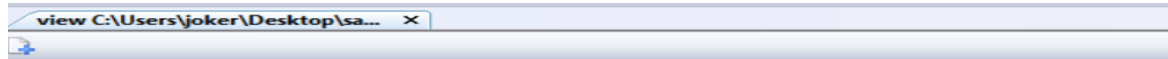
Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	412.8325	11.36821	22.73642	376.6538	449.0112
1	4	384.75	26.37045	52.74089	300.8275	468.6725
combined	8	398.7912	14.31339	40.48438	364.9455	432.637
diff		28.0825	28.71649		-42.18421	98.34921

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.8171
Ha: diff != 0
Pr(|T| > |t|) = 0.3659
Ha: diff > 0
Pr(T > t) = 0.1829
t = 0.9779
degrees of freedom = 6

```

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. ttest Firsthalfanhour , by (Animal)
Ready
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```
. ttest Firsthalfanhour , by (Animal)
Two-sample t test with equal variances
```

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	908.665	21.29462	42.58924	840.896	976.434
1	4	1077.418	16.75467	33.50935	1024.097	1130.738
combined	8	993.0412	34.26915	96.92779	912.0076	1074.075
diff		-168.7525	27.09575		-235.0534	-102.4516

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.0004
Ha: diff != 0
Pr(|T| > |t|) = 0.0008
Ha: diff > 0
Pr(T > t) = 0.9996
t = -6.2280
degrees of freedom = 6

```

```
. ttest secondhour , by (Animal)
Two-sample t test with equal variances
```

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	1153.25	9.440726	18.88145	1123.205	1183.295
1	4	1140	9.026248	18.0525	1111.274	1168.726
combined	8	1146.625	6.544259	18.50996	1131.15	1162.1
diff		13.25	13.06141		-18.71012	45.21012

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.8252
Ha: diff != 0
Pr(|T| > |t|) = 0.3495
Ha: diff > 0
Pr(T > t) = 0.1748
t = 1.0144
degrees of freedom = 6

```

```
. ttest Secondhalfanhour , by (Animal)
```

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. ttest Secondhalfanhour , by (Animal)

Two-sample t test with equal variances

```

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	1145.333	17.71451	35.42901	1088.957	1201.708
1	4	1125.75	16.92717	33.85433	1071.88	1179.62
combined	8	1135.541	11.93056	33.74472	1107.33	1163.753
diff		19.5825	24.50169		-40.37097	79.53597

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.7727
Ha: diff != 0
Pr(|T| > |t|) = 0.4546
Ha: diff > 0
Pr(T > t) = 0.2273
t = 0.7992
degrees of freedom = 6

. ttest Thirdhour , by (Animal)

Two-sample t test with equal variances

```

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	1119	14.23293	28.46586	1073.704	1164.296
1	4	1080.75	20.24246	40.48492	1016.329	1145.171
combined	8	1099.875	13.54498	38.31099	1067.846	1131.904
diff		38.25	24.74537		-22.29974	98.79974

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.9134
Ha: diff != 0
Pr(|T| > |t|) = 0.1731
Ha: diff > 0
Pr(T > t) = 0.0866
t = 1.5457
degrees of freedom = 6

. ttest Thirdhalfanhour , by (Animal)

```

```

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. ttest Thirdhalfanhour , by (Animal)

Two-sample t test with equal variances

```

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	1034.958	21.41065	42.82129	966.8193	1103.096
1	4	990.5425	6.74575	13.4915	969.0745	1012.01
combined	8	1012.75	13.35801	37.78215	981.1633	1044.337
diff		44.415	22.44818		-10.51373	99.34373

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.9524
Ha: diff != 0
Pr(|T| > |t|) = 0.0952
Ha: diff > 0
Pr(T > t) = 0.0476
t = 1.9786
degrees of freedom = 6

. import excel "C:\Users\joker\Downloads\goat-sheep.xlsx", sheet("Sheet4") firstrow clear

. ttest zerohour , by (Animal)

Two-sample t test with equal variances

```

Group	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
0	4	129.0825	3.78285	7.565701	117.0438	141.1212
1	4	131.415	14.32518	28.65036	85.82589	177.0041
combined	8	130.2488	6.872733	19.43902	113.9973	146.5002
diff		-2.3325	14.81623		-38.58651	33.92151

```

diff = mean(0) - mean(1)
Ho: diff = 0
Ha: diff < 0
Pr(T < t) = 0.4400
Ha: diff != 0
Pr(|T| > |t|) = 0.8801
Ha: diff > 0
Pr(T > t) = 0.5600
t = -0.1574
degrees of freedom = 6

```

```

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. ttest firsthour , by (Animal)

Two-sample t test with equal variances



| Group    | Obs | Mean     | Std. Err. | Std. Dev. | [95% Conf. Interval] |          |
|----------|-----|----------|-----------|-----------|----------------------|----------|
| 0        | 4   | 322.375  | 24.59276  | 49.18552  | 244.1099             | 400.6401 |
| 1        | 4   | 371.71   | 26.76551  | 53.53102  | 286.5302             | 456.8898 |
| combined | 8   | 347.0425 | 19.23642  | 54.40882  | 301.5556             | 392.5294 |
| diff     |     | -49.335  | 36.34827  |           | -138.276             | 39.606   |



diff = mean(0) - mean(1)
Ho: diff = 0
degrees of freedom = 6
t = -1.3573

Ha: diff < 0
Pr(T < t) = 0.1118

Ha: diff != 0
Pr(|T| > |t|) = 0.2235

Ha: diff > 0
Pr(T > t) = 0.8882

. ttest Firsthalfanhour , by (Animal)

Two-sample t test with equal variances



| Group    | Obs | Mean      | Std. Err. | Std. Dev. | [95% Conf. Interval] |           |
|----------|-----|-----------|-----------|-----------|----------------------|-----------|
| 0        | 4   | 856.9575  | 20.39206  | 40.78411  | 792.0609             | 921.8541  |
| 1        | 4   | 1009      | 13.73105  | 27.4621   | 965.3017             | 1052.698  |
| combined | 8   | 932.9787  | 30.90492  | 87.4123   | 859.9002             | 1006.057  |
| diff     |     | -152.0425 | 24.58409  |           | -212.1976            | -91.88739 |



diff = mean(0) - mean(1)
Ho: diff = 0
degrees of freedom = 6
t = -6.1846

Ha: diff < 0
Pr(T < t) = 0.0004

Ha: diff != 0
Pr(|T| > |t|) = 0.0008

Ha: diff > 0
Pr(T > t) = 0.9996

. ttest secondhour , by (Animal)

```

```

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. ttest secondhour , by (Animal)

Two-sample t test with equal variances



| Group    | Obs | Mean     | Std. Err. | Std. Dev. | [95% Conf. Interval] |          |
|----------|-----|----------|-----------|-----------|----------------------|----------|
| 0        | 4   | 1063.083 | 21.37557  | 42.75115  | 995.0559             | 1131.109 |
| 1        | 4   | 1121.5   | 22.55038  | 45.10076  | 1049.735             | 1193.265 |
| combined | 8   | 1092.291 | 18.13167  | 51.28412  | 1049.417             | 1135.166 |
| diff     |     | -58.4175 | 31.07145  |           | -134.4466            | 17.61159 |



diff = mean(0) - mean(1)
Ho: diff = 0
degrees of freedom = 6
t = -1.8801

Ha: diff < 0
Pr(T < t) = 0.0546

Ha: diff != 0
Pr(|T| > |t|) = 0.1091

Ha: diff > 0
Pr(T > t) = 0.9454

. ttest Secondhalfanhour , by (Animal)

Two-sample t test with equal variances



| Group    | Obs | Mean     | Std. Err. | Std. Dev. | [95% Conf. Interval] |          |
|----------|-----|----------|-----------|-----------|----------------------|----------|
| 0        | 4   | 1167.5   | 24.51734  | 49.03467  | 1089.475             | 1245.525 |
| 1        | 4   | 1186.747 | 31.19053  | 62.38106  | 1087.485             | 1286.01  |
| combined | 8   | 1177.124 | 18.7218   | 52.95324  | 1132.854             | 1221.394 |
| diff     |     | -19.2475 | 39.67303  |           | -116.3239            | 77.8289  |



diff = mean(0) - mean(1)
Ho: diff = 0
degrees of freedom = 6
t = -0.4852

Ha: diff < 0
Pr(T < t) = 0.3224

Ha: diff != 0
Pr(|T| > |t|) = 0.6448

Ha: diff > 0
Pr(T > t) = 0.6776

. ttest Thirdhour , by (Animal)

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. ttest Thirdhour , by (Animal)

Two-sample t test with equal variances

+-----+-----+-----+-----+-----+-----+
| Group | Obs | Mean | Std. Err. | Std. Dev. | [95% Conf. Interval] |
+-----+-----+-----+-----+-----+-----+
| 0      | 4   | 1076.168 | 27.05919 | 54.11837 | 990.0531 1162.282 |
| 1      | 4   | 1076.918 | 16.7328  | 33.4656  | 1023.666 1130.169 |
+-----+-----+-----+-----+-----+-----+
| combined | 8   | 1076.543 | 14.72811 | 41.65738 | 1041.716 1111.369 |
+-----+-----+-----+-----+-----+-----+
| diff    |     | -.75    | 31.81487 |           | -78.59819 77.09819 |
+-----+-----+-----+-----+-----+-----+
diff = mean(0) - mean(1)
Ho: diff = 0
degrees of freedom = 6
t = -0.0236

Ha: diff < 0      Ha: diff != 0      Ha: diff > 0
Pr(T < t) = 0.4910  Pr(|T| > |t|) = 0.9820  Pr(T > t) = 0.5090

. ttest Thirdhalfanhour , by (Animal)

Two-sample t test with equal variances

+-----+-----+-----+-----+-----+-----+
| Group | Obs | Mean | Std. Err. | Std. Dev. | [95% Conf. Interval] |
+-----+-----+-----+-----+-----+-----+
| 0      | 4   | 935.25 | 22.34623 | 44.69247 | 864.1343 1006.366 |
| 1      | 4   | 940.3325 | 21.18483 | 42.36967 | 872.9129 1007.752 |
+-----+-----+-----+-----+-----+-----+
| combined | 8   | 937.7912 | 14.28628 | 40.40771 | 904.0096 971.5729 |
+-----+-----+-----+-----+-----+-----+
| diff    |     | -5.0825 | 30.79207 |           | -80.42797 70.26297 |
+-----+-----+-----+-----+-----+-----+
diff = mean(0) - mean(1)
Ho: diff = 0
degrees of freedom = 6
t = -0.1651

Ha: diff < 0      Ha: diff != 0      Ha: diff > 0
Pr(T < t) = 0.4372  Pr(|T| > |t|) = 0.8743  Pr(T > t) = 0.5628

. exit, clear
ready

```

BIOGRAPHY

I am Md. Sayem Abdullah, son of Abu Ahmed & Momtaz Begum. I passed my Secondary School Certificate examination from Bandarban Govt. High School in 2014 & Higher Secondary Certificate examination from Bandarban Cantonment Public School & College in 2016. I enrolled for Doctor of Veterinary Medicine (DVM) degree in Chattogram Veterinary and Animal Sciences University (CVASU), Bangladesh. In future, I would like to work as a veterinary practitioner.